

Using Dirichlet Gaussian Processes to Analyze Gene Expression of Cancer Metastasis Progression

Perla Molina
BEEHIVE Lab
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Introduction

It's me!



Perla Molina

- First Year PhD in DBDS
- Bachelor's in Data Science at USF
 - DaVita Internship
 - AWM President
- Why Stanford?
 - Easy move
 - Meaningful research
 - Data science realm
- Research interests
 - Cancer and disease
 - gynecology/women's health
- Obsessed with kpop and horror movies



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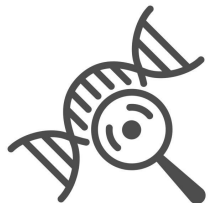
Background Info + Material



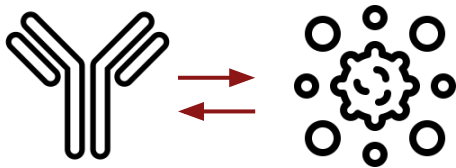
The Biological Problem



Cancer metastasis is the cause of death for **50-90%** → no current therapies to specifically target metastasis [2] → the need to look at metastatic data



Understanding genetic dynamics of cancer metastasis remains incomplete [2] → lots of undiscovered territories, especially with progression over time



Previous computational analysis reveals “an ordered series of immunological changes that correspond to metastatic progression” [2] → importance of looking at differential changes at molecular and genetic levels → potential for target-based therapies



What is DP_GP?

- Bayesian nonparametric model for time series trajectories [1]
 - P is the number of genes
 - T the number of time points per sample, assuming observations at the same time points across samples, but allowing for missing observations (missing data)
- Bayesian part → probabilistic framework that can analyze uncertainty
- “**DP clusters the trajectories** of gene expression levels across time, where the **trajectories are modeled using a Gaussian process.**” [1]

$$Y \in \mathbb{R}^{P \times T}$$

$$G \sim DP(\alpha, G_0);$$

$$\theta_h \sim G;$$

$$y_j \sim p(\cdot | \theta_h).$$



Why DP_GP?



Benefit: Do not have to assume the given number of clusters at beginning, a priori [1] (other methods mostly do) → huge benefit for analyzing differential growth over time



Benefit: Does not assume independence of clusters (like k-Means, hierarchical clustering, etc) → important in clustering gene expression over period of time

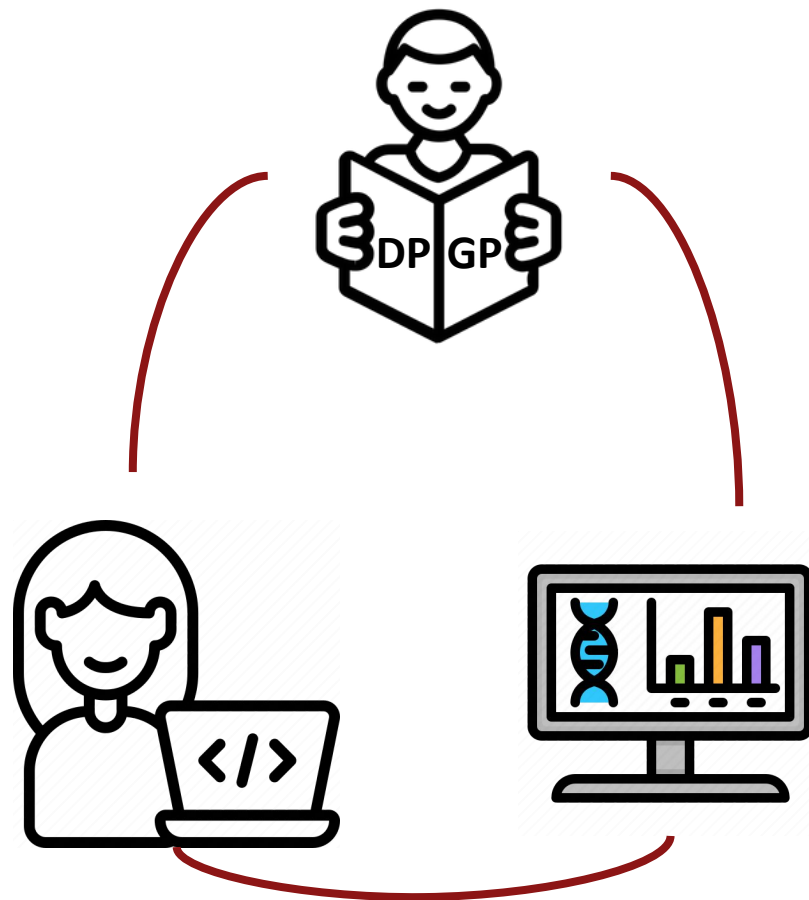


Objectives



My Task

- Learn how DP_GP works
- Update and install DP_GP software
- Extract, preprocess, and format data
 - Look at top 3 and lowest 2 frequent cell types
- Analyze gene expression of metastatic lung cancer in mice over time using DP_GP

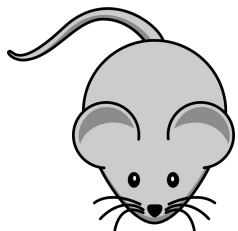


Methodology



Study Design

29 mice total



Sequence cells
over 14 weeks

Read genes



Wet lab ———
Me ———

Read counts

	Cell Type 1 - Wk0Rep1	Cell Type 1 - Wk1Rep2	Cell Type 2 - Wk6Rep1	...
Gene 1				
Gene 2				
Gene 3				
Gene 4				
...				

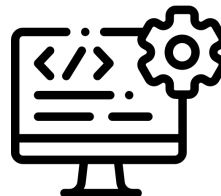
Data Extraction



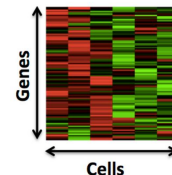
Preprocessing



Run DP_GP



Analyze gene expression



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What I Used



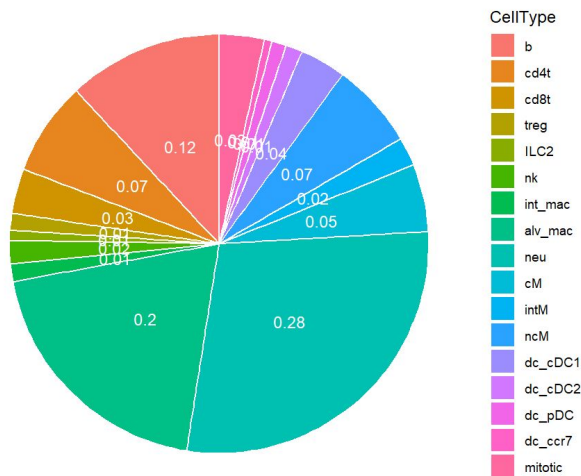
- R
 - Extract data for each cell type
 - Format by individual timepoints
 - Sum counts of each replicate & timepoint
 - Preprocess & select significant genes
 - CPM value threshold ≥ 10 [3]
 - Log2 fold change threshold ≥ 4 and adjusted Wilcox p-value < 0.01 [4][5]
 - Bonferroni correction
 - Normalize significant genes
 - Average the sums of replicates for each time point
 - Z-score normalization
- Python
 - Fix & update outdated code
 - Install updated DP_GP
 - Run DP_GP on final output data from R (45 iterations per cell type dataset)



Results

Top & Lowest Frequent Cell Types

Cell Type by Frequency %



Top 3

CellType	Freq
neu	24017
alv_mac	16593
b	10111

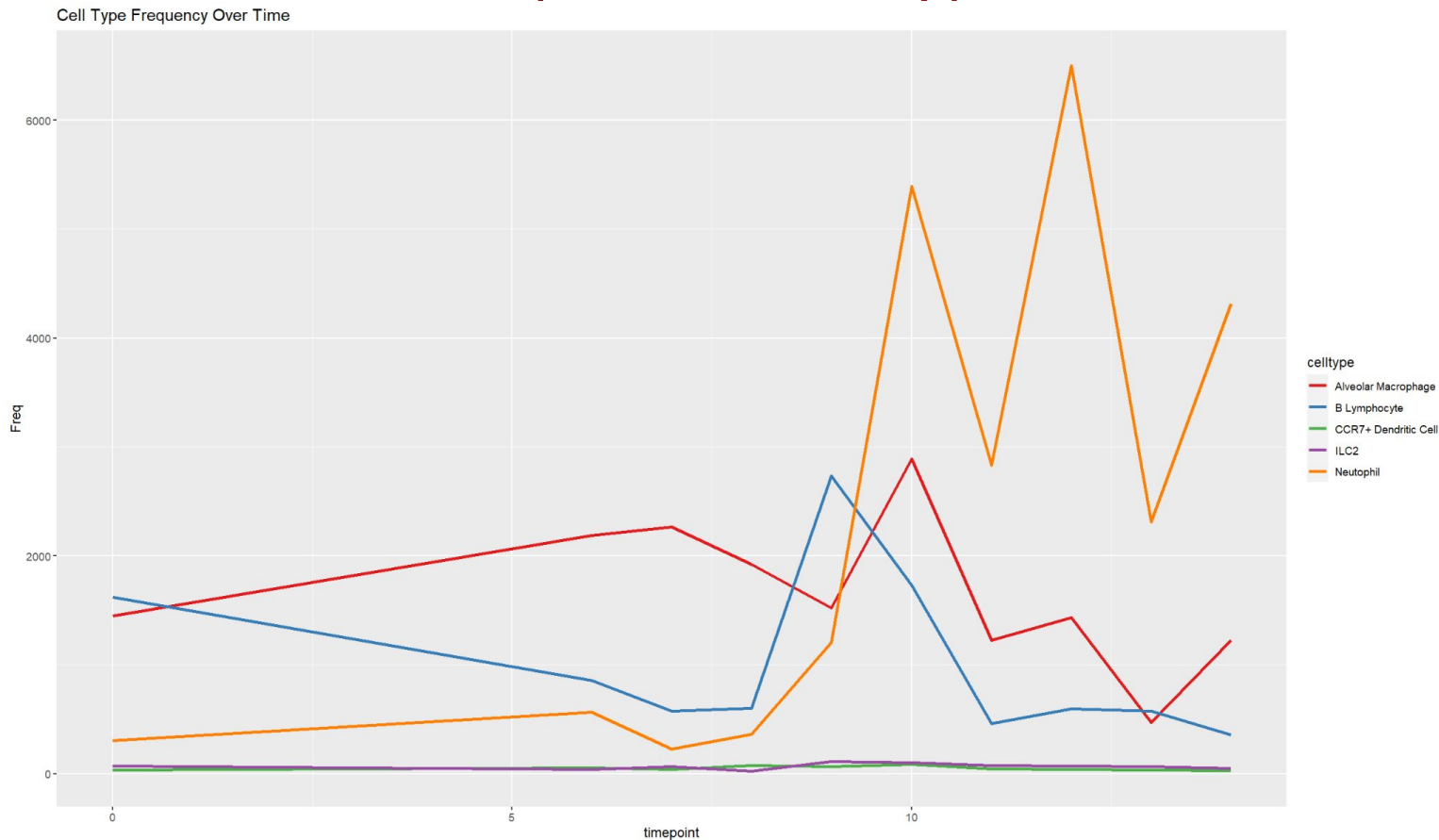
Lowest 2

CellType	Freq
ILC2	683
dc_ccr7	505

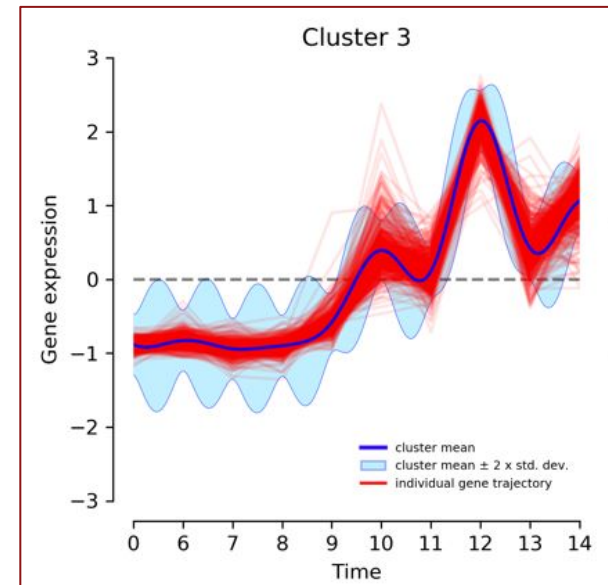
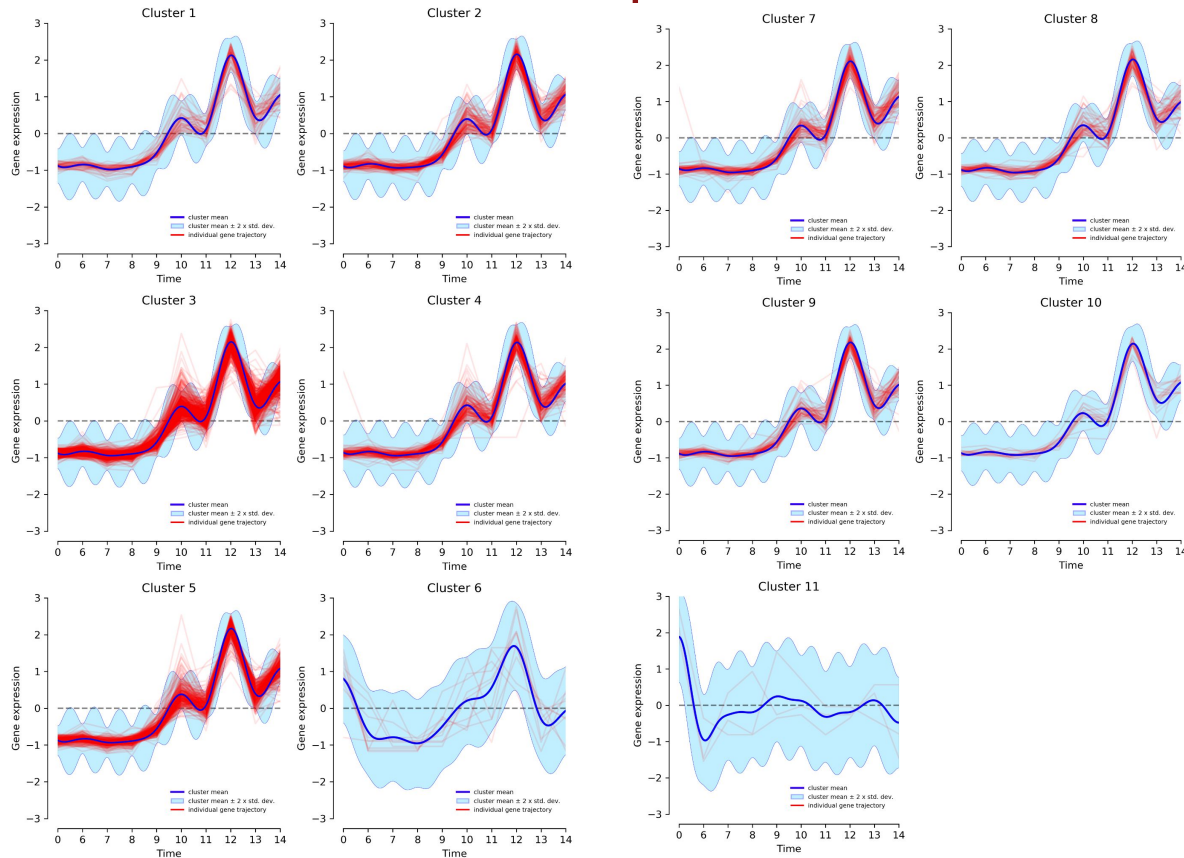
- neu = Neutrophil cells
- alv_mac = Alveolar macrophages
- b = B lymphocyte cells
- ILC2 = Type 2 Innate Lymphoid Cells
- dc_ccr7 = CCR7+ Dendritic cells



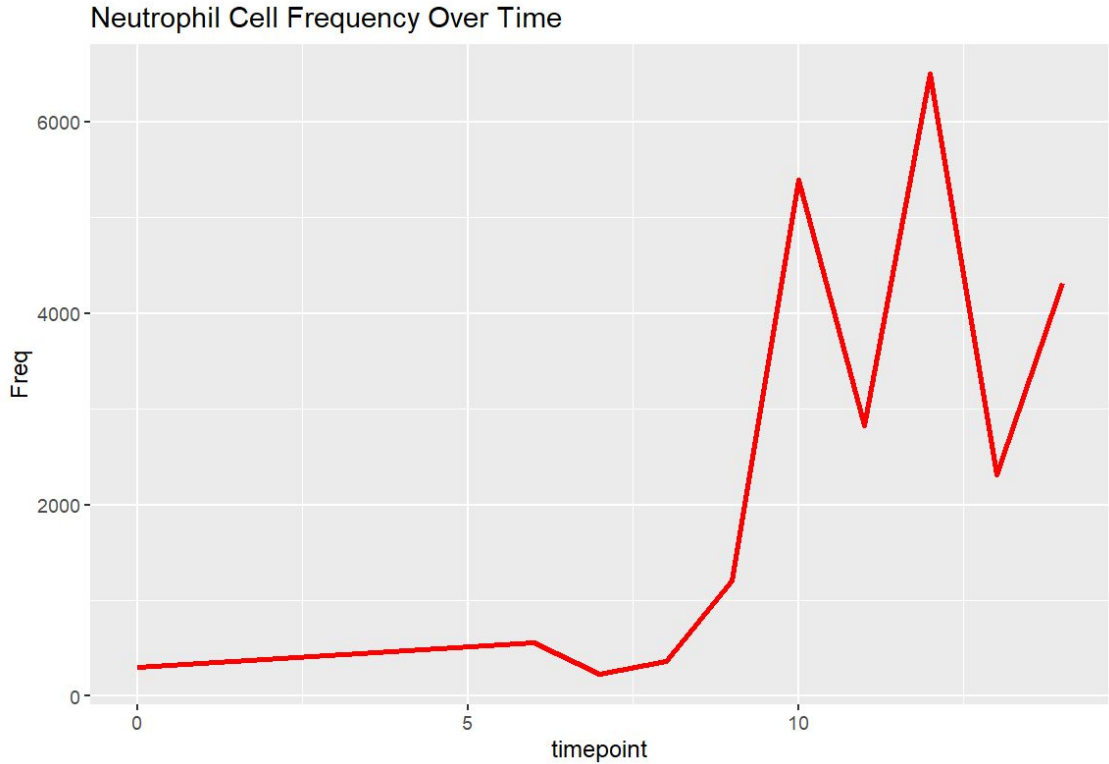
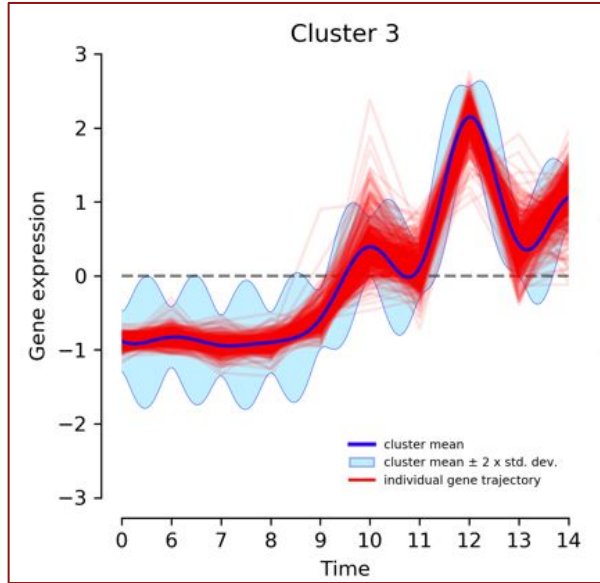
Top & Lowest Frequent Cell Types Over Time



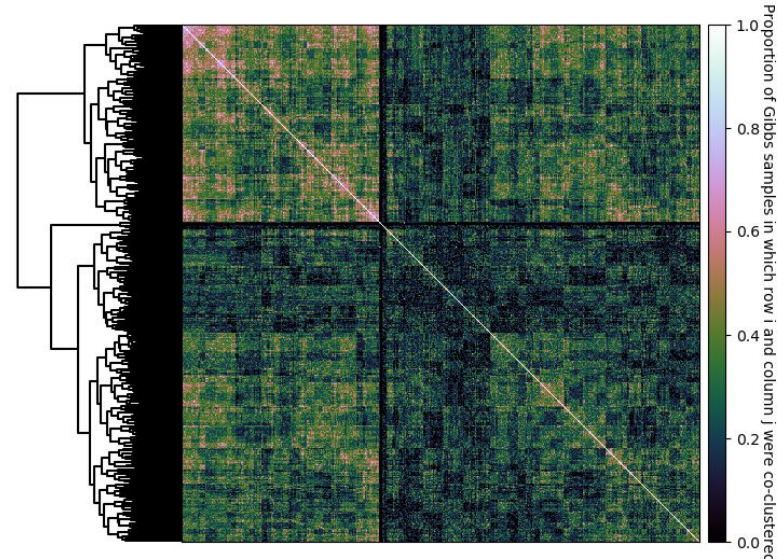
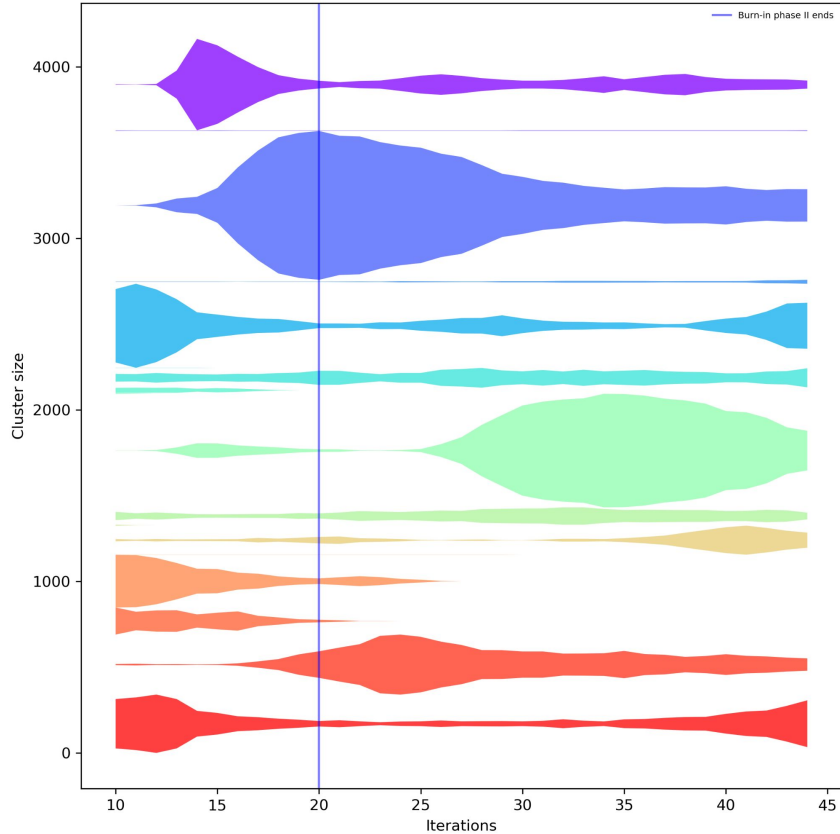
DP_GP Gene Expression for Neutrophil Cells



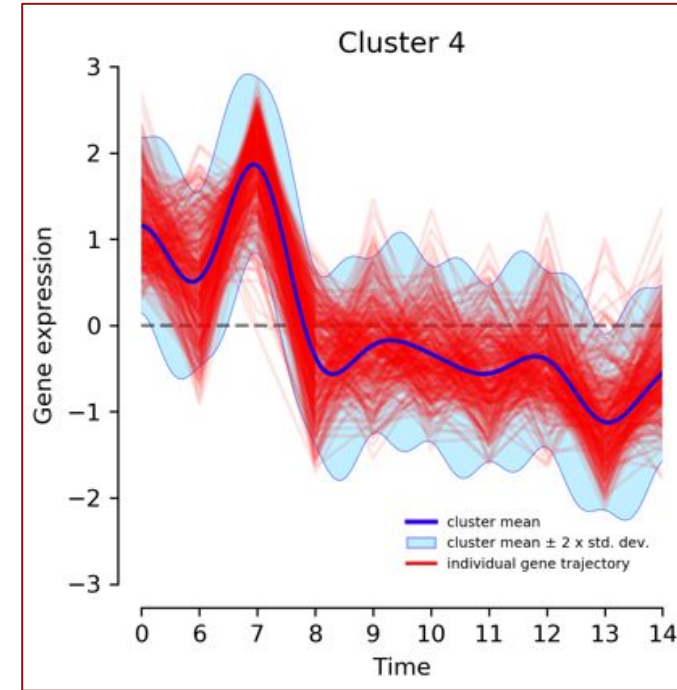
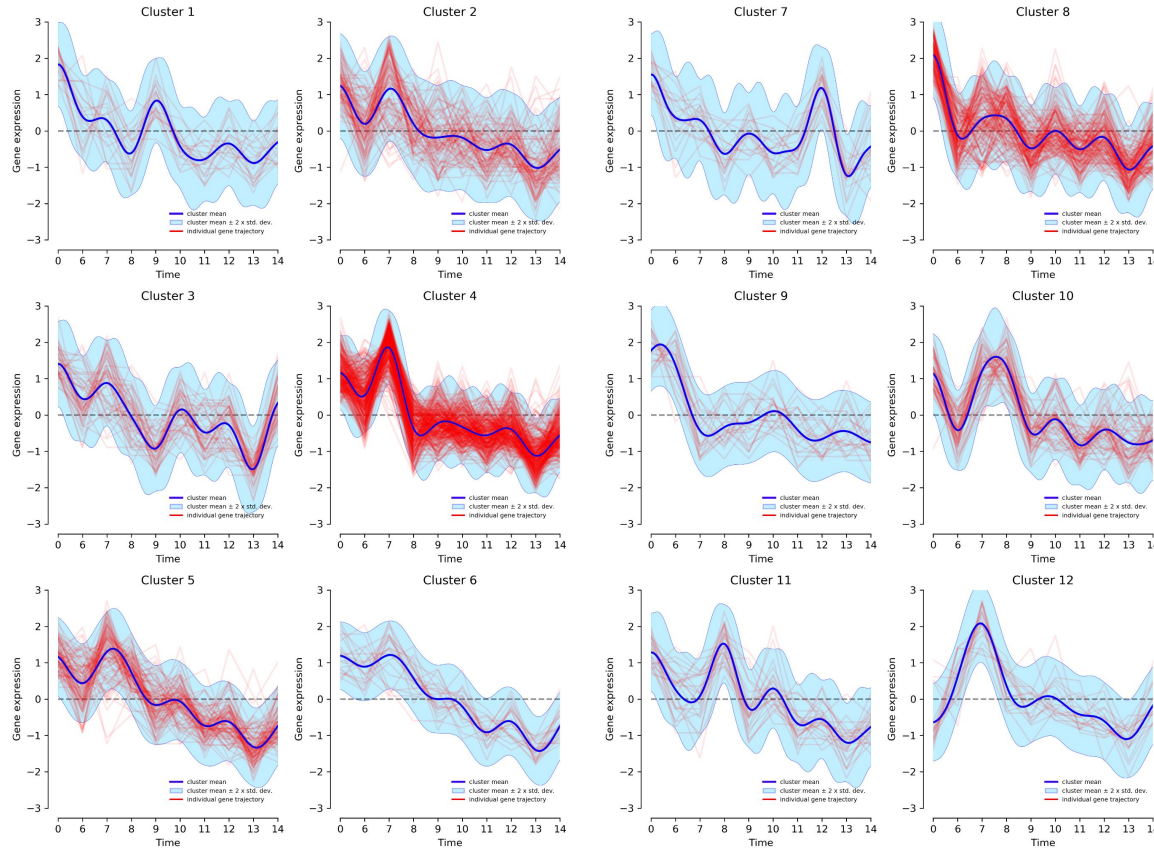
Closer Look at Neutrophil Cells Over Time



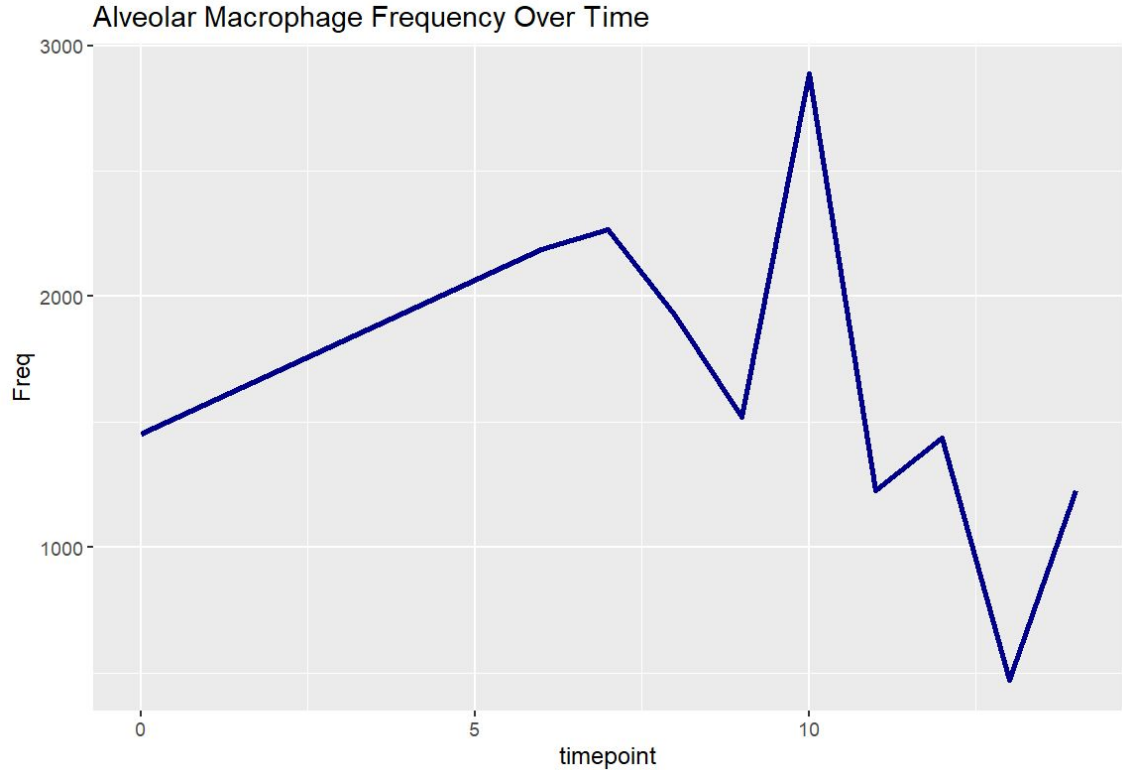
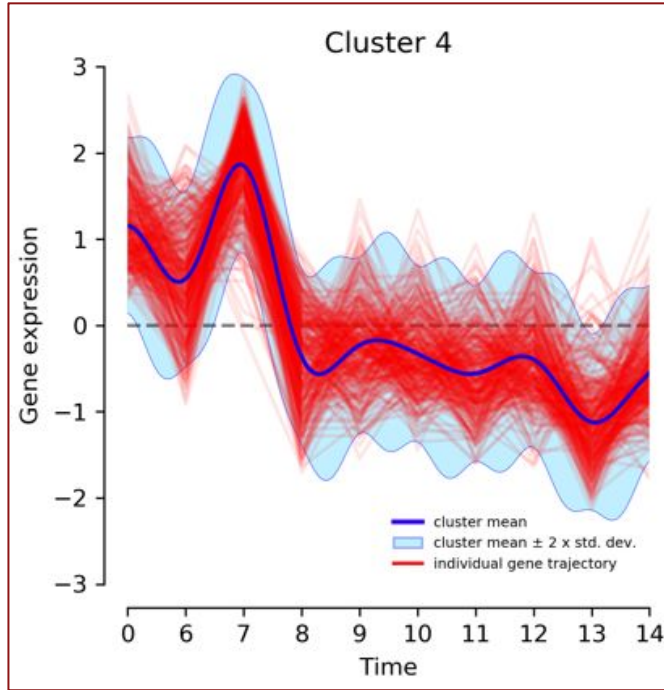
DP_GP Iteration Results for Neutrophil Cells



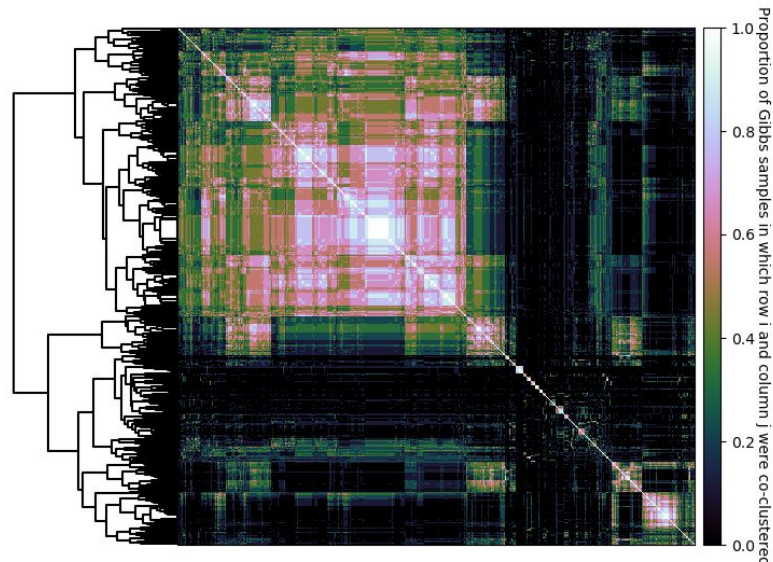
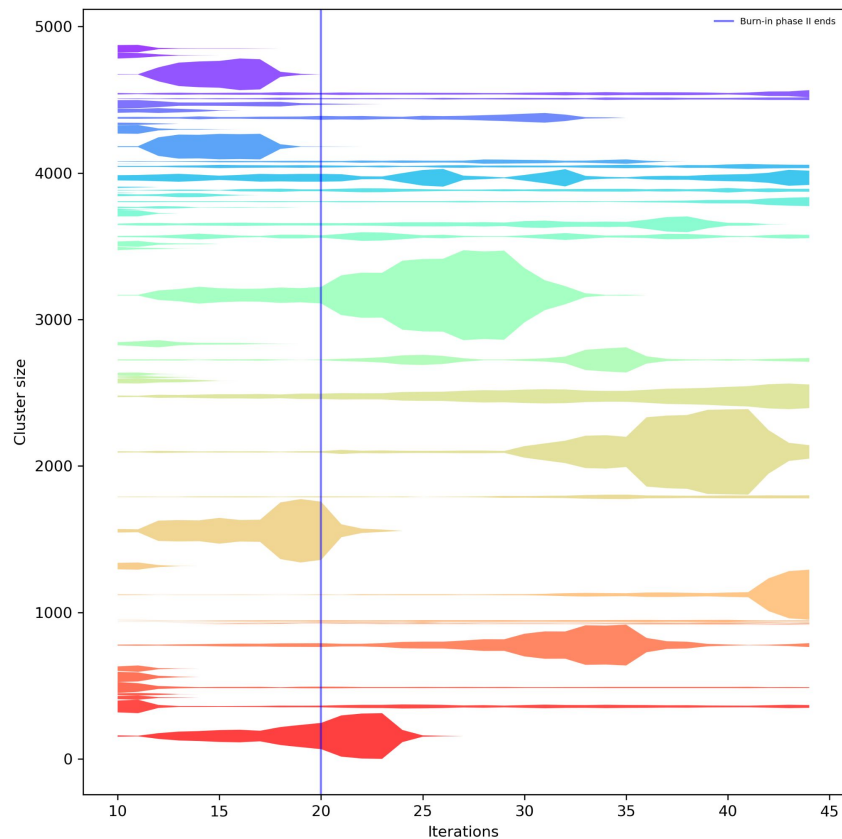
DP_GP Gene Expression for Alveolar Macrophages



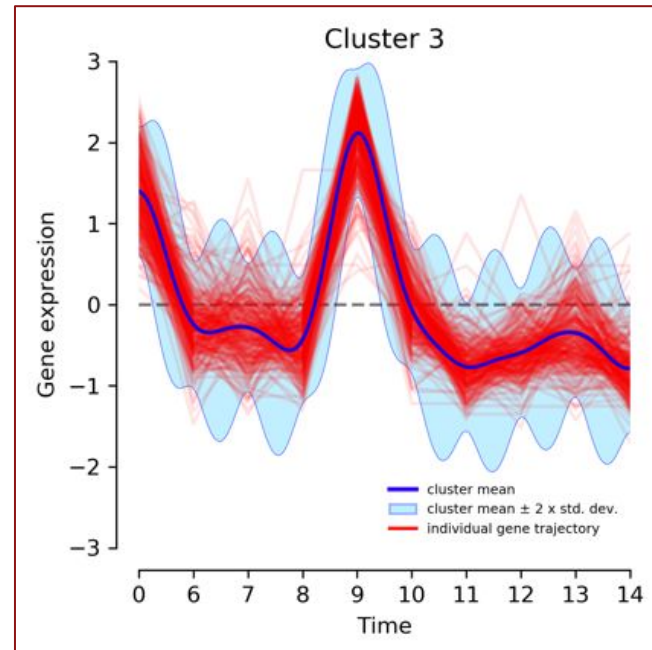
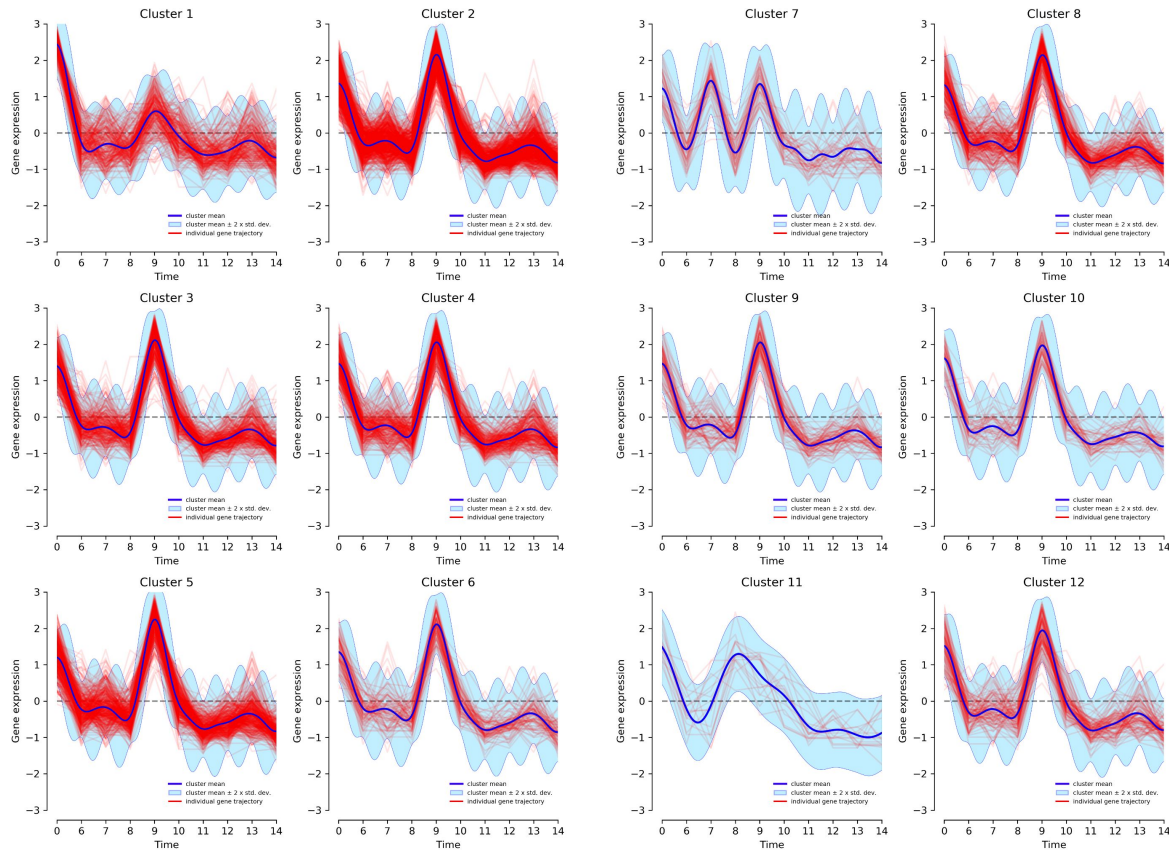
Closer Look at Alveolar Macrophages Over Time



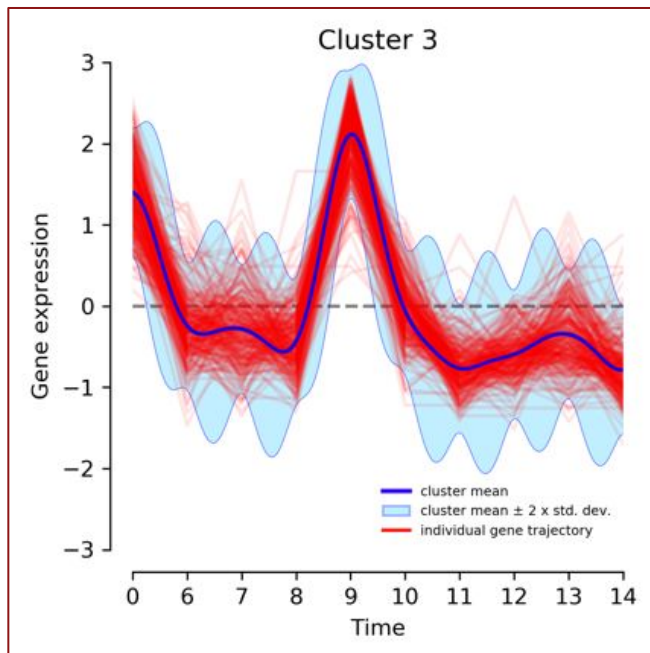
DP_GP Iteration Results for Alveolar Macrophages



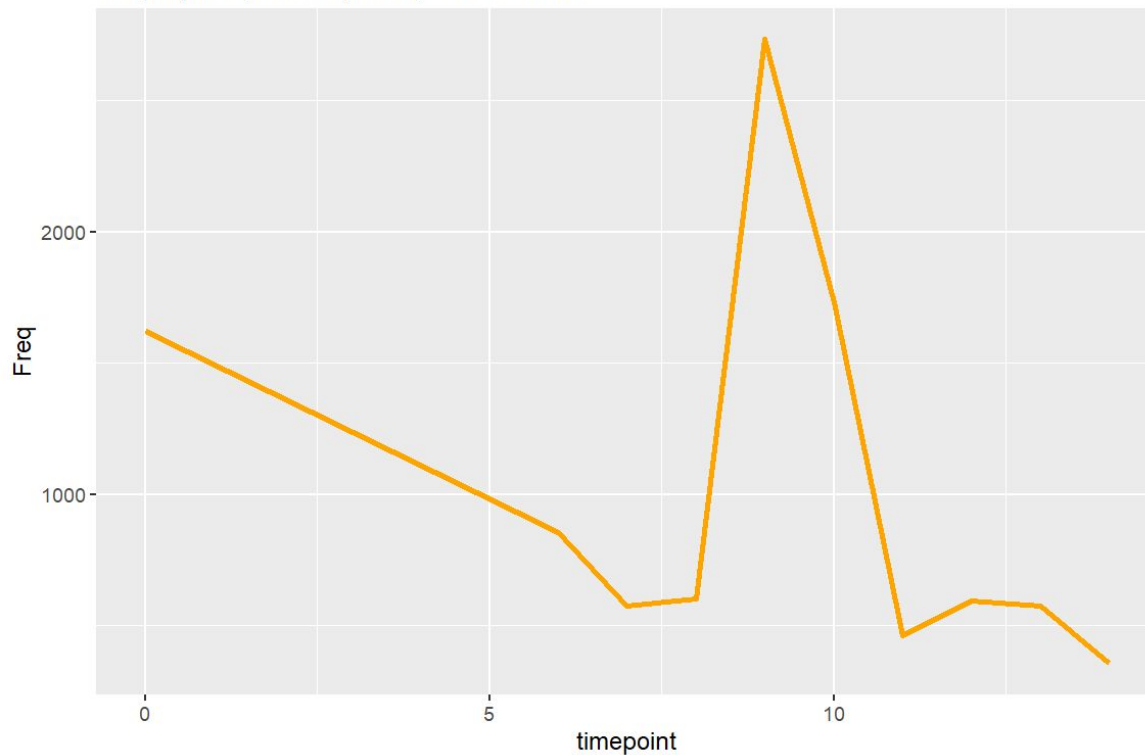
DP_GP Gene Expression for B Lymphocytes



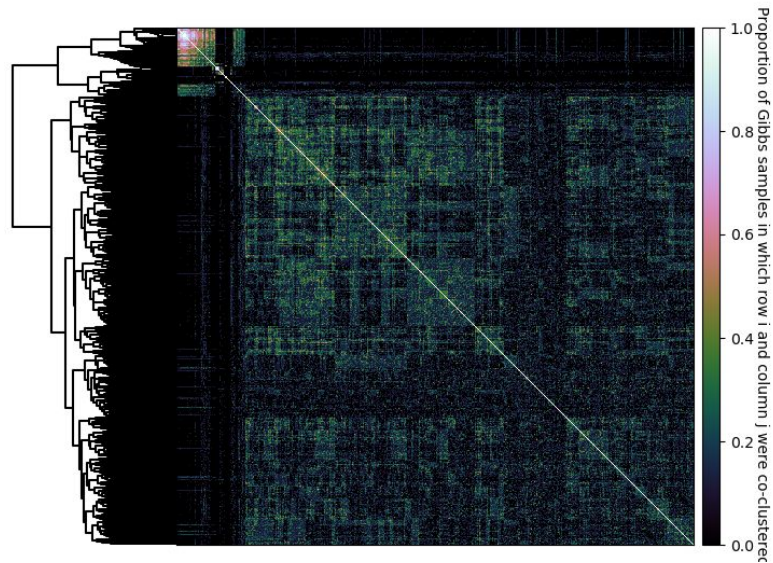
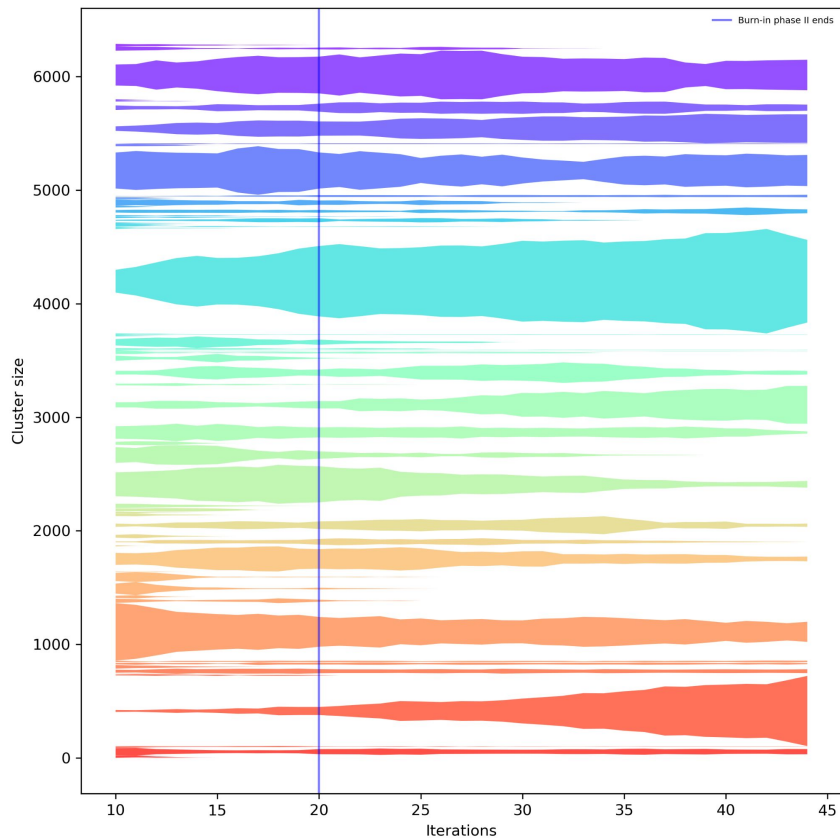
Closer Look at B Lymphocytes



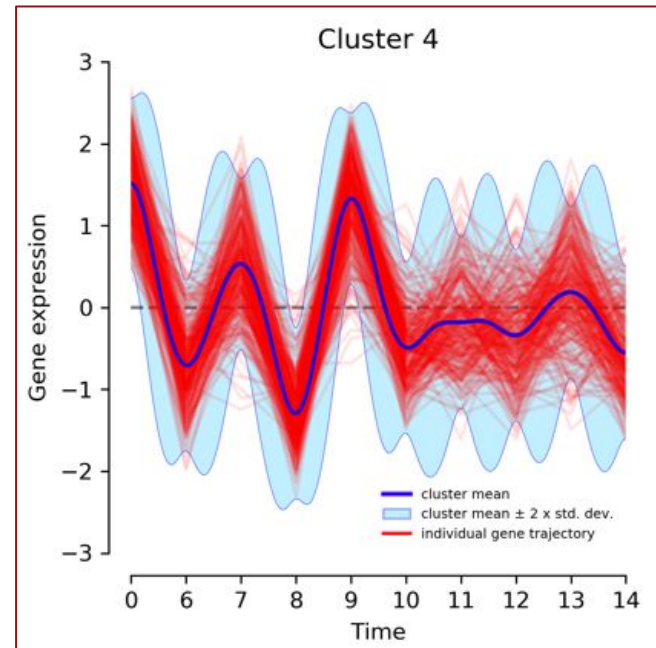
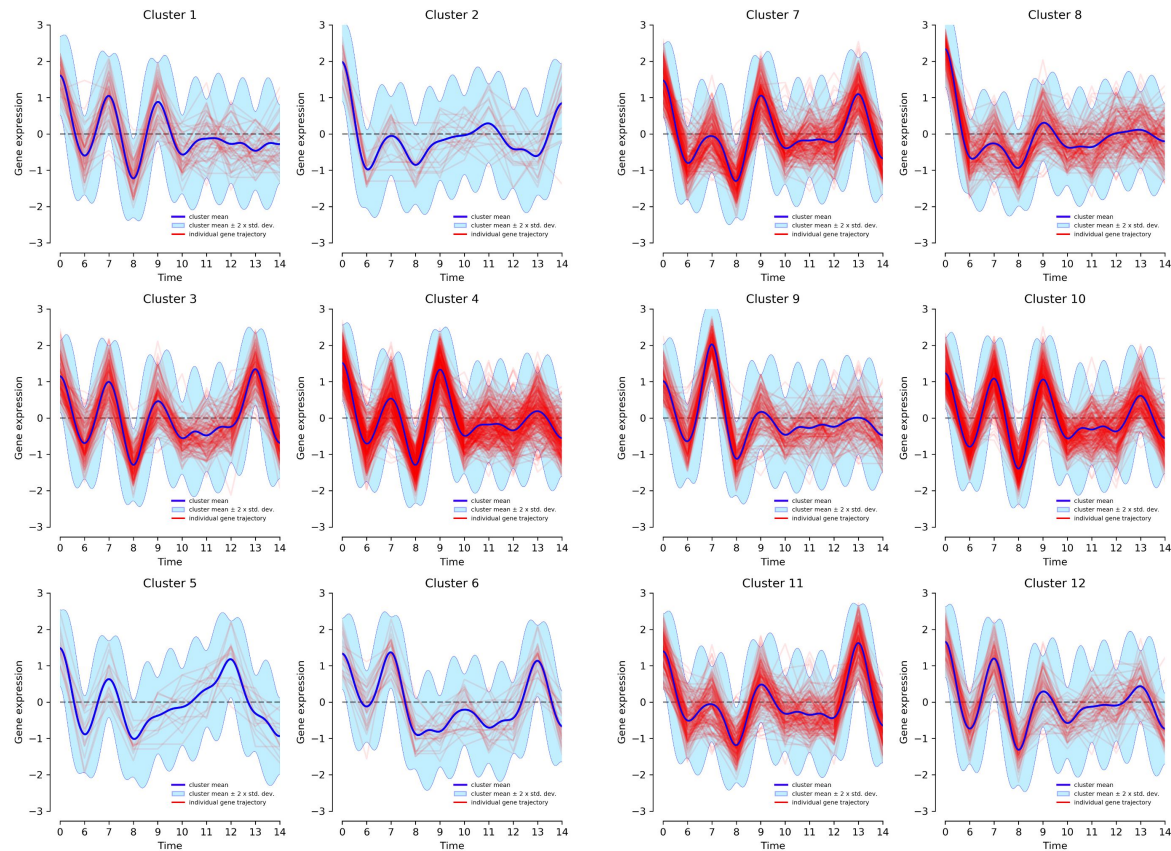
B Lymphocyte Frequency Over Time



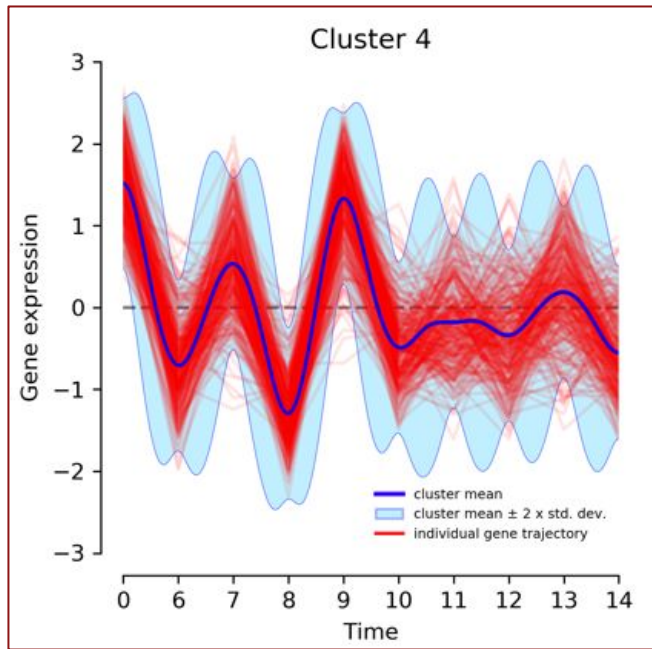
DP_GP Iteration Results for B Lymphocytes



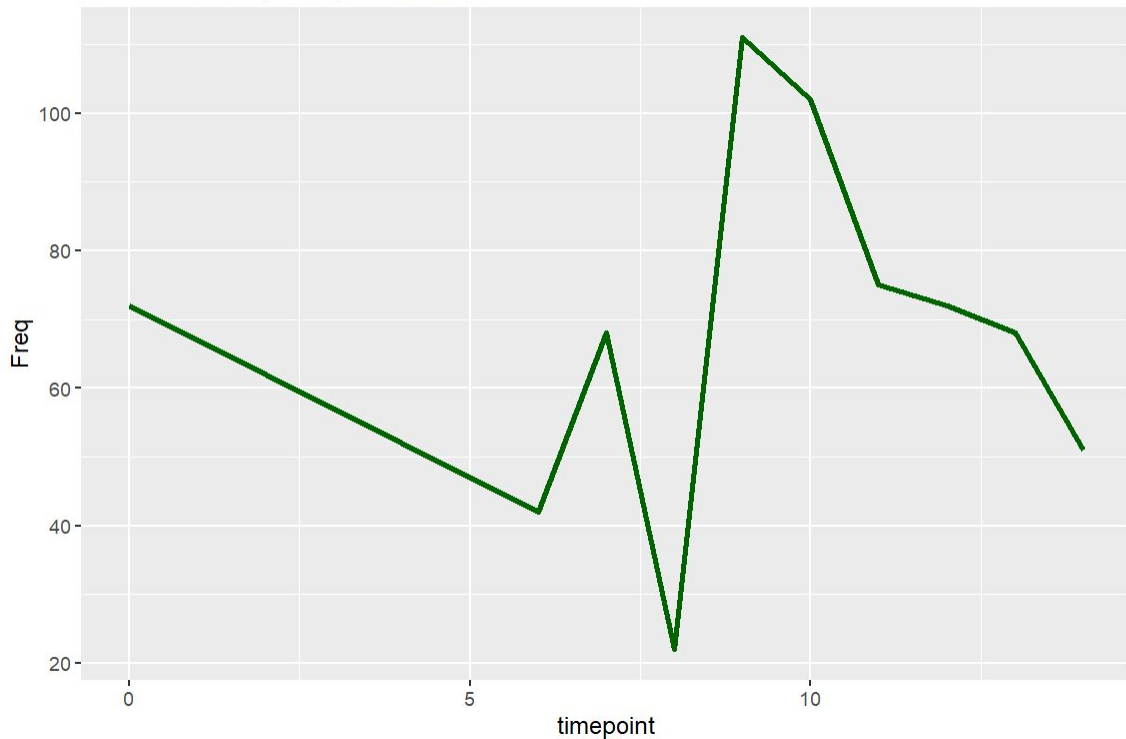
DP_GP Gene Expression for Type 2 Innate Lymphoid Cells



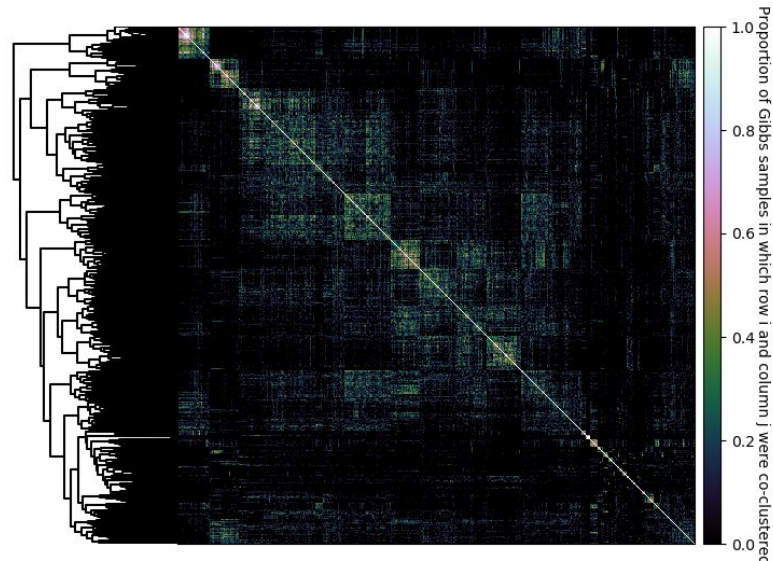
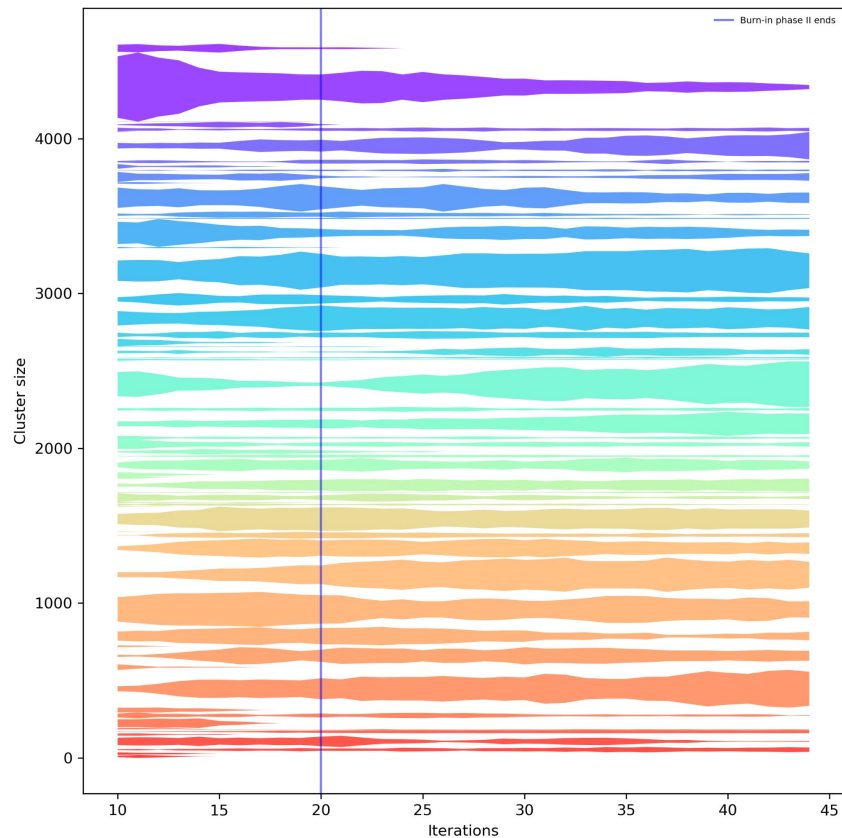
Closer Look at Type 2 Innate Lymphoid Cells



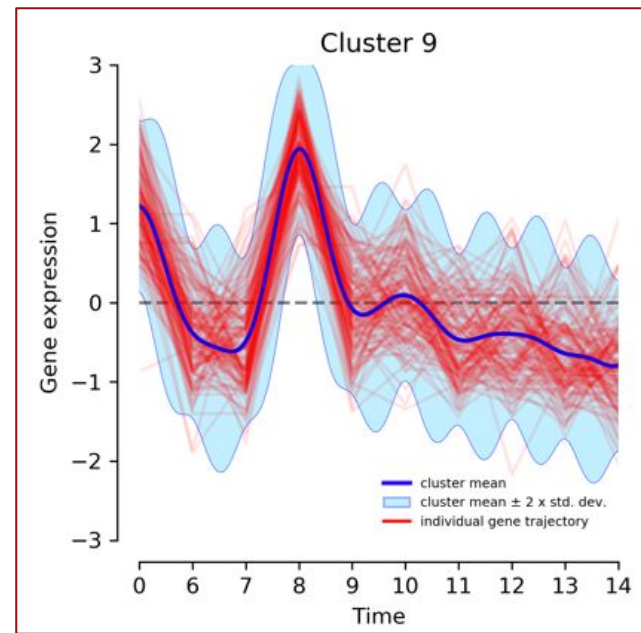
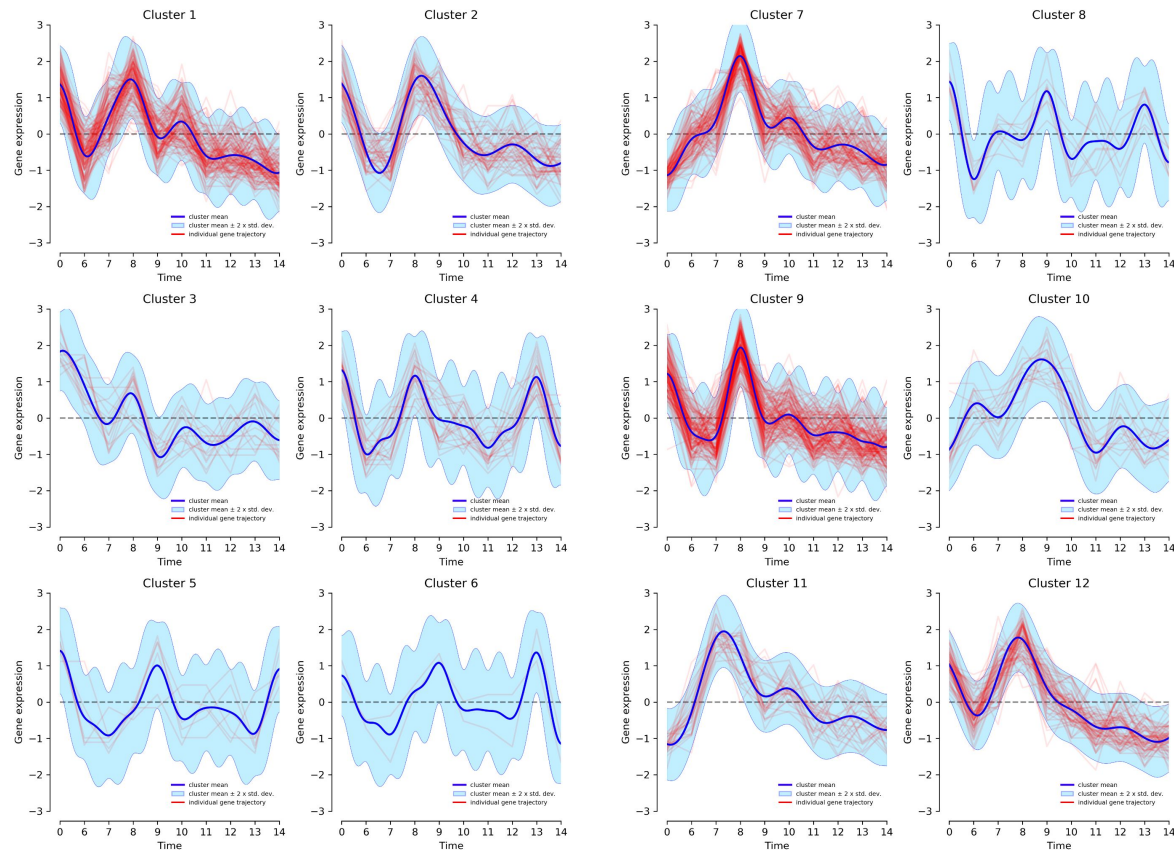
ILC2 Cell Frequency Over Time



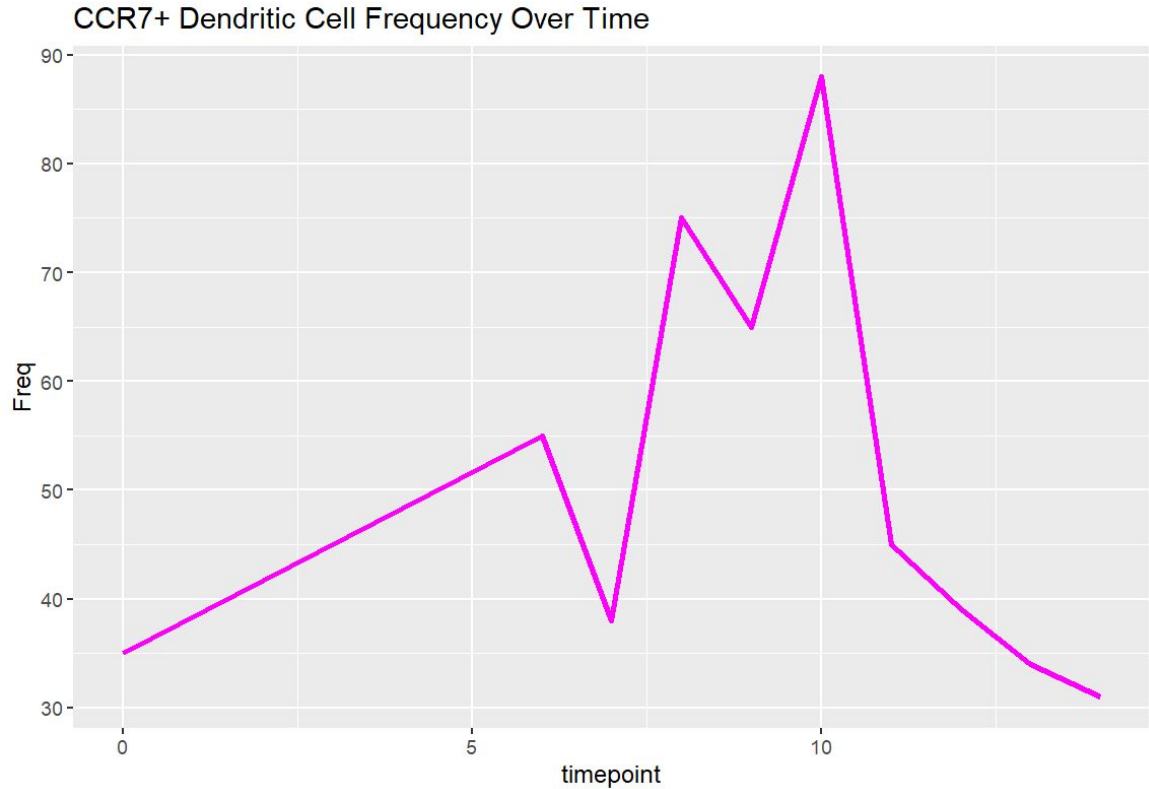
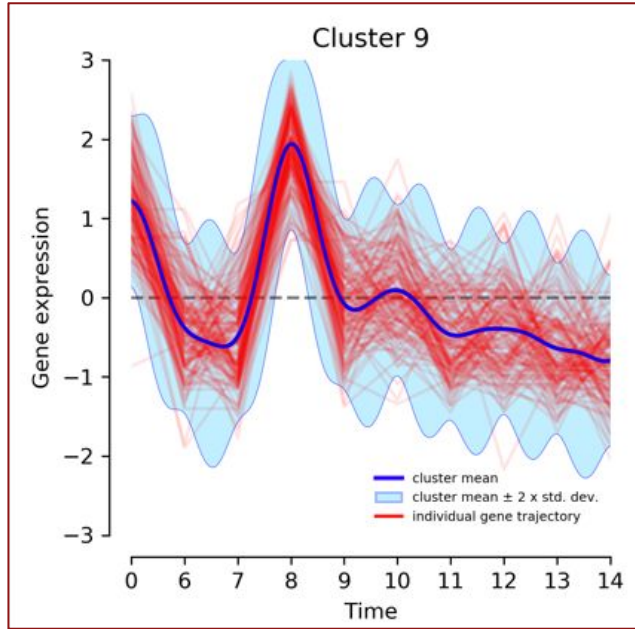
DP_GP Iteration Results for Type 2 Innate Lymphoid Cells



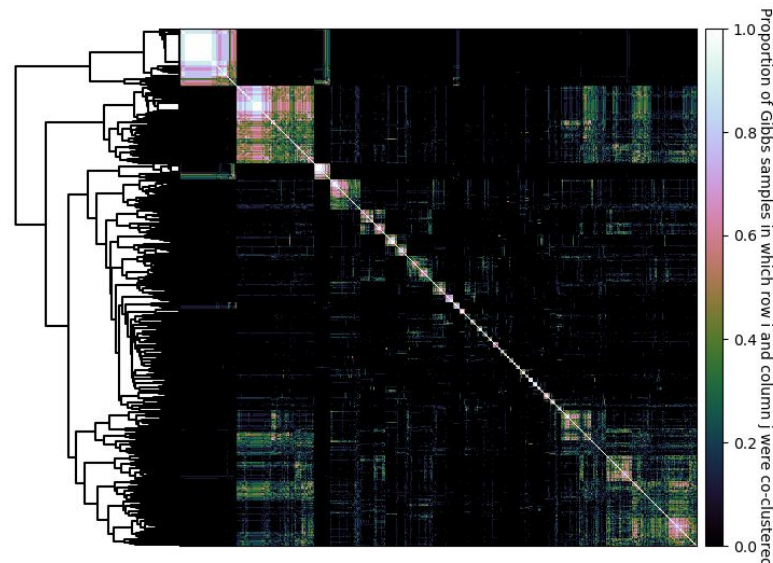
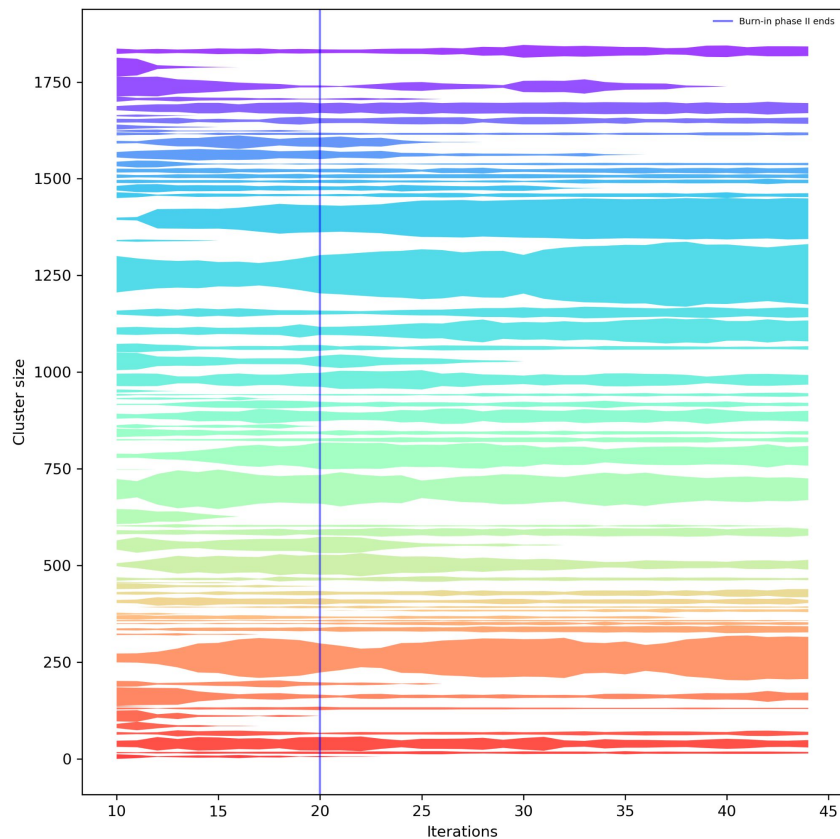
DP_GP Gene Expression for CCR7+ Dendritic Cells



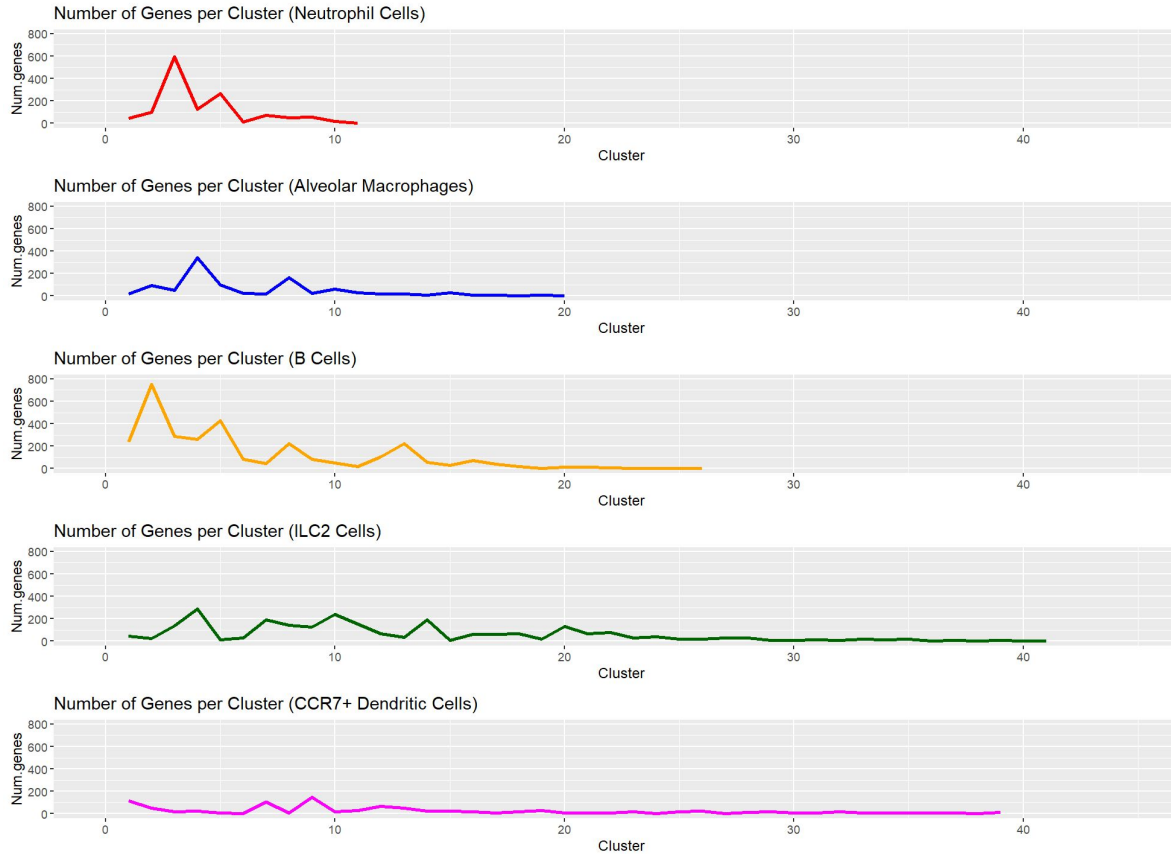
Closer Look at CCR7+ Dendritic Cells



DP_GP Iteration Results for CCR7+ Dendritic Cells



Benefit of DP_GP Clustering



Conclusions + Implications



Key Conclusions + (Possible) Implications

Conclusion	Implication
Correlations between gene expression and number of cells over time	More cells increases gene expression → connections between up-down regulation and frequency of cell type → potential for target-based therapies and indications for dosages
Most frequent cells were related to immunity/immune-response functions	The body does try its best to combat cancer, but still didn't fully perform its job → potential for target-based therapies & research for cell-cell interactions in cancer metastasis
Key spikes of gene expression at certain time points	Relativity to the progression of illness (i.e. extreme gene expression (up or down-reg) correspond to specific cycles or pathways of cancer) → potential for outcomes research (pinpoint time of cancer progression that is the worst/least worst) OR unknown issue during data collection in lab



Key Conclusions + (Possible) Implications

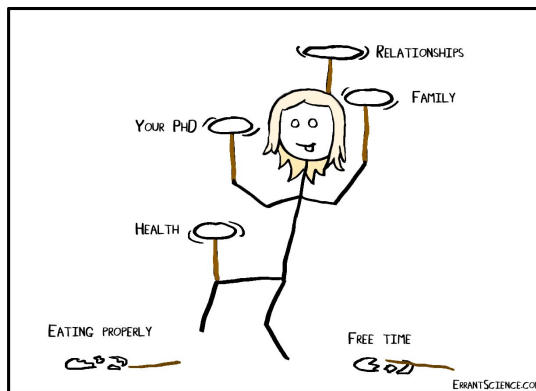
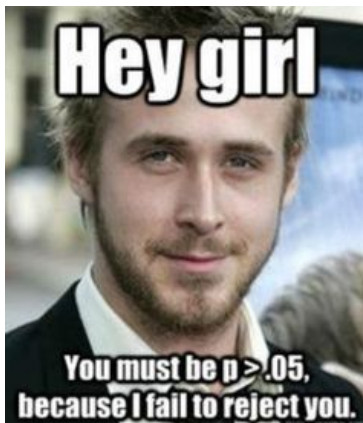
Conclusion	Implication
Number of clusters increased with the less frequent cell types	Less frequent clusters have extreme variability and uncertainty → rare cell types or possible “by-stander” cells in cancer progression → less certain about their purpose and importance → optimization problems and possible indication of cancer progression having less impact on gene expression for these specific cell types
Overall optimal number of clusters $\sim <10$	DP_GP will need to be iterated more to condense clusters

Challenges & Opportunities



Challenges

- Updating DP_GP code
- Extracting and formatting data
- Figuring out the best statistical thresholds
- Adjusting to grad school life right after undergrad (balancing classes and research)



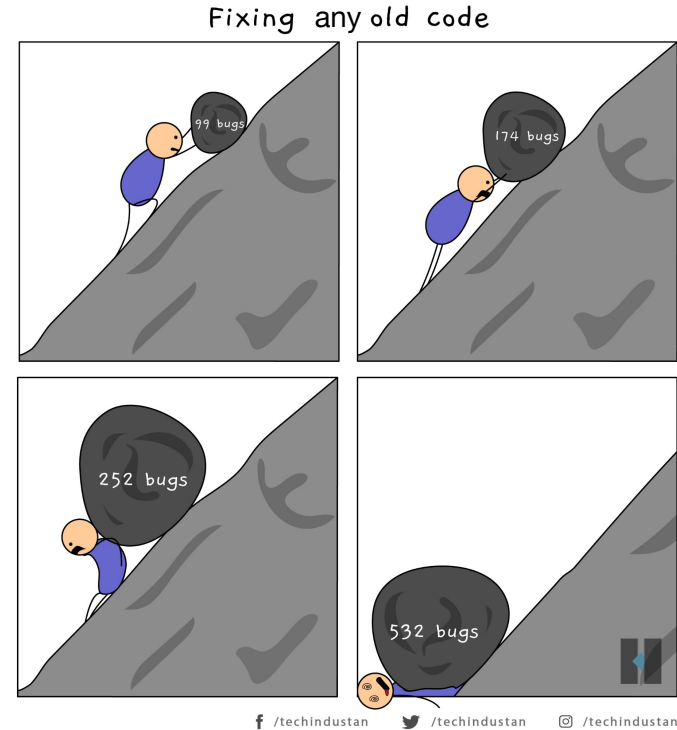
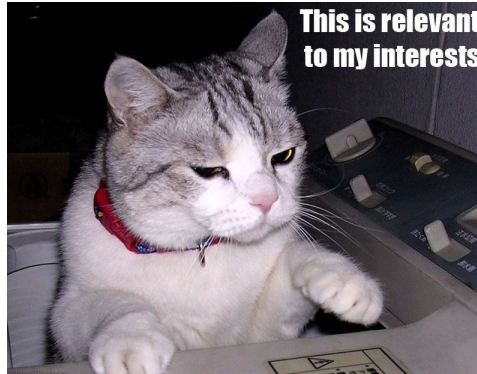
TRYING TO DO EVERYTHING DURING
A PHD CAN BE A BIT TRICKY



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Opportunities (What I Learned)

- The importance of making user-friendly and up-to-date code/software/methods
- The endless possibilities of research in gene expression alone and the various different ways to analyze it
- Developed an interest in RNA-seq analysis/research



If I Had More Time

- Analyze all cell types from the data (17 total)
- Combat the memory issues to run more iterations → better clusters
- Run a DP_GP analysis on immune-response cells vs non-immune-response
- Find a way to collectively run DP_GP on the entire dataset (not just individual cell types)
- Conduct cross-validation techniques or methods for gene expression results
- Improve DP_GP software to efficiently work with single cell data (initially used to analyze bacterial growth)



References

References

- [1] McDowell IC, Manandhar D, Vockley CM, Schmid AK, Reddy TE, et al. (2018) Clustering gene expression time series data using an infinite Gaussian process mixture model. PLOS Computational Biology 14(1): e1005896.
<https://doi.org/10.1371/journal.pcbi.1005896>.
- [2] McGinnis, Christopher S., et al. “The Temporal Progression of Immune Remodeling during Metastasis.” bioRxiv, Cold Spring Harbor Laboratory, 1 Jan. 2023, www.biorxiv.org/content/10.1101/2023.05.04.539153v1.
- [3] Law, Charity. RNA-Seqbasics: From Reads to Differential Expression - Github Pages, combine-australia.github.io/RNAseq-R/slides/RNASeq_basics.pdf.
- [4] McCarthy DJ, Smyth GK. Testing significance relative to a fold-change threshold is a TREAT. Bioinformatics. 2009 Mar 15;25(6):765-71. doi: 10.1093/bioinformatics/btp053. Epub 2009 Jan 28. PMID: 19176553; PMCID: PMC2654802.
- [5] “Lab #5 Differential Expression.” Data Analysis, www.bioconductor.org/help/course-materials/2015/Uruguay2015/day5-data_analysis.html.

Q&A + Acknowledgements

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